

## An approach to heterobifunctional poly(ethyleneglycol) bioconjugates

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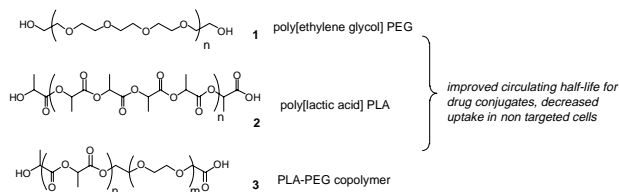
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**Abstract**—A family of differentially substituted poly(ethyleneglycol) building blocks has been assembled from commercially available material. Their utility is demonstrated by formation of amino acid conjugates, image contrast agents, gold nanoparticles, and functional antibody conjugates. Application in the cellular trafficking of antitumoral agent conjugates is expected.  
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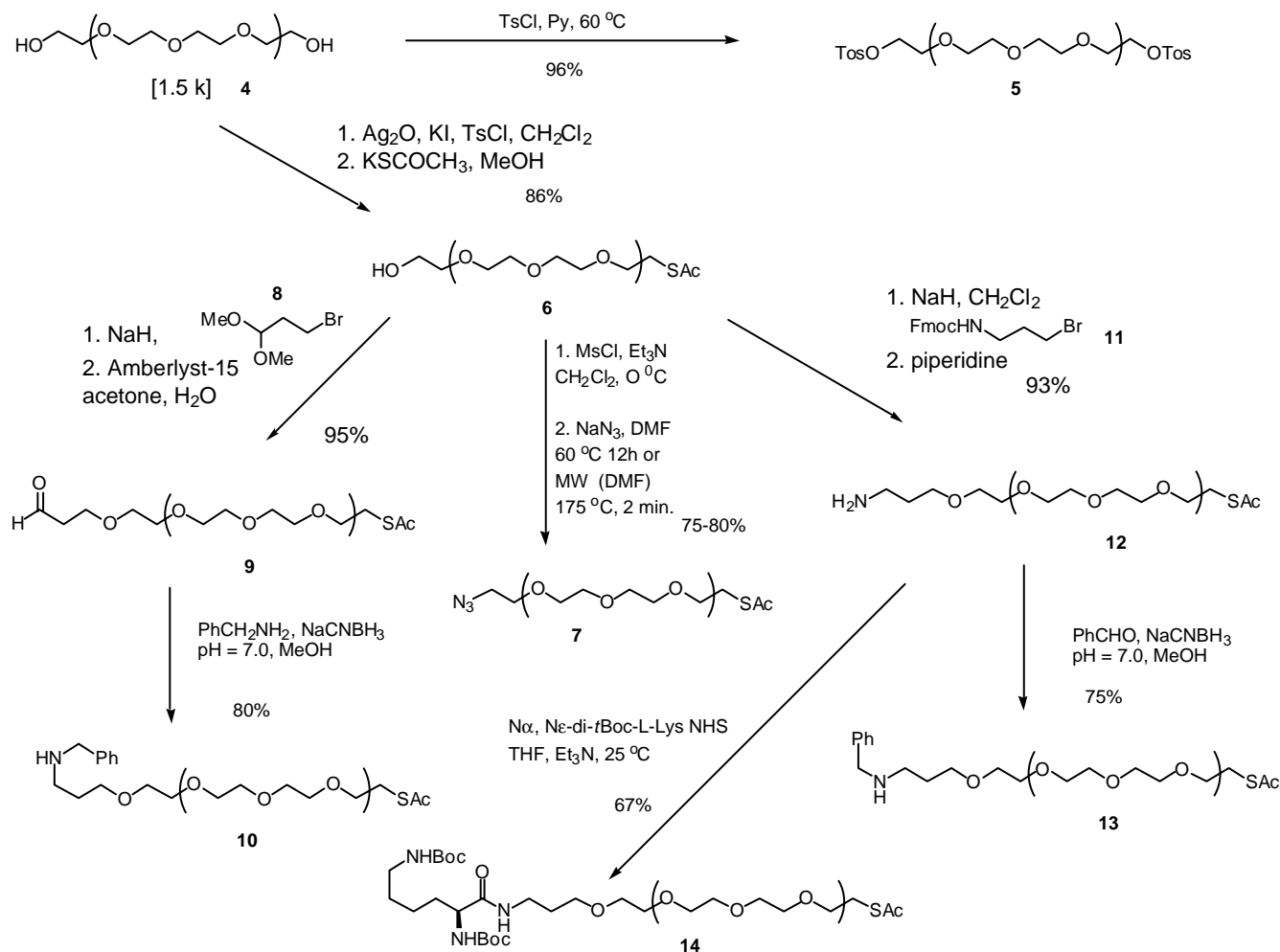
One of the goals of targeted drug delivery is to enhance uptake at the desired site of action using the minimum effective dose of agent.<sup>1</sup> In the case of many solid tumors, it has been shown that derivatizing or encapsulating drugs with polymers including poly(ethyleneglycols) **1** can greatly improve their circulating half-lives and in some cases enhance uptake in tumors by exploiting the leaky vasculature.<sup>2</sup> This strategy can also help overcome immunogenic reactions to protein based drugs, and is currently being employed in a clinical trial of an enediyne–chromoprotein complex, with encapsulation in a block-copolymer.<sup>3</sup> In the field of prostate cancer, liposome encapsulated drugs have been explored as have polymer coupled agents. One of these, a poly(ethyleneglycol) (PEG) linked doxorubicin, has shown enhanced efficacy.<sup>4</sup> While PEG derivatives

continue to be evaluated,<sup>2</sup> there is considerable interest in related polymers including lactic acids (**2**) and hybrids, e.g., the PEG–polylactic acids (PLA, **3**), a nanoparticle conjugate of which has been shown to adhere to prostate membrane specific antigen (PMSA) expressing cells when coupled to aptamers that bind PMSA.<sup>5</sup> Accordingly, there is considerable interest in derivatives of reagent-grade polymers, which can be readily tailored for specific purposes, e.g., attachment of aptamers, fluorescent imaging agents, antibodies, and motifs to enhance nuclear uptake.<sup>6</sup>

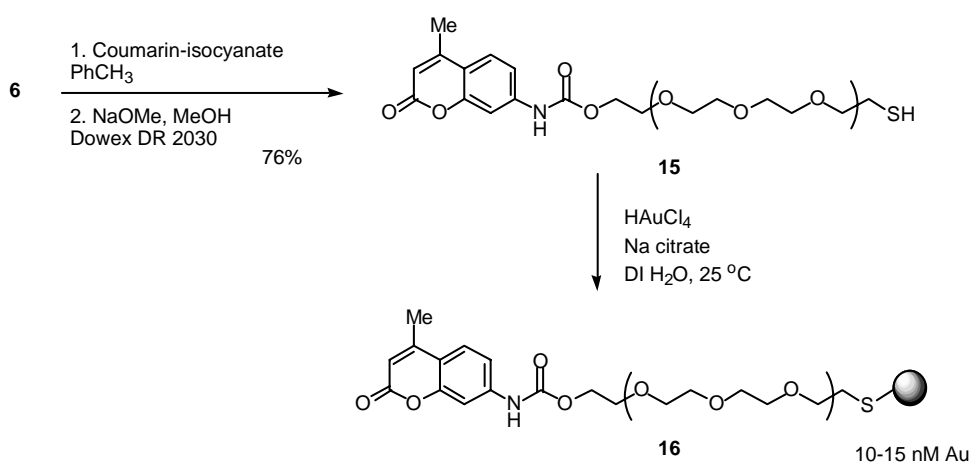
Our initial objective was to demonstrate a route to bifunctional building blocks commencing with commercially available starting materials. An uncapped 1.5K average molecular weight PEG diol **4** (Aldrich) was selected and subjected to desymmetrization (Scheme 1). Bis tosylation to **5** can be avoided by use of silver oxide/KI, and the resulting monotosylate was converted to thioacetate **6** via nucleophilic displacement with the potassium salt.<sup>7</sup> Compound **6** proved highly versatile for production of numerous derivatives, the thio group selected on the basis of potential addition to surfaces using established thiol chemistry.<sup>8</sup> Conversion to the corresponding mesylate of **6** proved more efficient than the tosylate, allowing azido displacement to give **7**, which may have application as a photoaffinity tag. Interestingly, microwave displacement (CEM Navigator system) was highly efficient giving an 80% yield of product



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Scheme 1. Preparation of bifunctional PEG building blocks.



Scheme 2. Preparation of coumarin and (Au) nanoparticle conjugates.

within 2 min. To introduce carboxaldehyde functionality, addition of bromoacetal **8** to the alkoxide salt of **6** was conducted and then the aldehyde was unmasked using Amberlyst resin. Product **9** was subjected to specimen reductive amination, benzylamine giving amine **10**

in good yield without compromise of the thioacetate group. Despite numerous efforts, attempted Staudinger reduction of azide **7** was unsuccessful,<sup>9</sup> thus we sought alternate means to introduce amine functionality. Remedy was found by alkylation with bromide **11**, which in



## References and notes

1. Langer, R. *Science* **2001**, 293, 5.
2. Roberts, M. J.; Bentley, M. D.; Harris, J. M. *Adv. Drug Delivery Rev.* **2002**, 54, 459.
3. Tsuchiya, K.; Uchida, T.; Kobayashi, M.; Maeda, H.; Konno, T.; Yamanaka, H. *Urology* **2000**, 55, 495.
4. Vaage, J.; Barbera, E. *Am. Assoc. Cancer Res.* **1994**, 35, 2481.
5. Farokhzad, O. C.; Jon, S.; Khademhosseini, A.; Tran, T.-N. T.; LaVan, D. A.; Langer, R. *Cancer Res.* **2004**, 64, 7668.
6. Aronov, O.; Horowitz, A. T.; Gabizon, A.; Fuertes, M. A.; Perez, J. M.; Gibson, D. *Bioconjugate Chem.* **2004**, 15, 814.
7. Bouzide, A.; Sauve, G. *Org. Lett.* **2002**, 4, 2329.
8. Characterizing spectroscopic and analytical data were obtained for all new compounds: **(7)**  $^1\text{H}$  NMR ( $\text{CDCl}_3$ , 300 MHz)  $\delta$  3.87 (m, 2H), 3.83–3.60 (m, 58H), 3.41 (m, 2H), 3.09 (m, 2H), 2.07 (s, 3H); **(10)**  $^1\text{H}$  NMR ( $\text{CDCl}_3$ , 300 MHz)  $\delta$  7.42–7.31 (m, 5H), 4.22 (m, 2H), 4.01–3.56 (m, 62H), 3.49 (m, 2H), 2.98–2.81 (m, 4H), 2.08 (s, 3H); **(13)**  $^1\text{H}$  NMR ( $\text{CDCl}_3$ , 300 MHz)  $\delta$  7.56–7.41 (m, 5H), 4.20 (m, 2H), 4.00–3.52 (m, 60H), 3.40 (m, 2H), 2.98 (m, 2H), 2.90–2.81 (m, 4H), 2.45 (br s, 1H), 2.08 (s, 3H); **(14)**  $^1\text{H}$  NMR ( $\text{CDCl}_3$ , 300 MHz)  $\delta$  8.0 (br s, 3H), 4.71 (m, 1H), 4.12–3.40 (m, 58H), 3.40–3.00 (m, 6H), 2.06 (s, 3H), 1.60–1.10 (m, 22H); **(15)**  $^1\text{H}$  NMR ( $\text{H}_2\text{O}$ , 300 MHz)  $\delta$  8.0 (br s, 1H), 7.52–7.42 (m, 3H), 6.17 (s, 1H), 4.35 (m, 2H), 3.94–3.63 (m, 58H), 3.50 (m, 2H), 2.83 (m, 2H), 2.41 (s, 3H).
9. Bertozzi, C. R.; Bednarski, M. D. *J. Org. Chem.* **1991**, 56, 4326.
10. Brust, M.; Walker, M.; Bethell, D.; Schiffrin, D. J.; Whyman, R. *J. Chem. Soc. Chem. Commun.* **1994**, 801; Brust, M.; Fink, J.; Bethell, D.; Schiffrin, D. J.; Kiely, C. *J. Chem. Soc. Chem. Commun.* **1995**, 1655; Rowe, M. P.; Plass, K. E.; Kim, K.; Kurdak, C.; Zellers, E. T.; Matzger, A. *J. Chem. Mater.* **2004**, 16, 3513; Kanaras, A. G.; Kamounah, F. S.; Schaumburg, K.; Kiely, C. J.; Brust, M. *J. Chem. Soc. Chem. Commun.* **2002**, 2294; Wuelfing, W. P.; Gross, S. M.; Miles, D. T.; Murray, R. W. *J. Am. Chem. Soc.* **1998**, 120, 12696; Mrksich, M.; Chen, C. S.; Xia, Y.; Dike, L. E.; Ingber, D. E.; Whitesides, G. M. *Proc. Natl. Acad. Sci. U.S.A.* **1996**, 93, 10775.
11. Determinations made using Coulter N4 sub-micron particle size analyzer. Nanoparticles were air-dried and specimen grid was observed with JEOL JEM-1010 TEM (voltage 60 kV, 250,000 $\times$  magnification).
12. Larson, R. S.; Menard, V.; Jacobs, H.; Kim, S. W. *Bioconjugate Chem.* **2001**, 12, 861; Chamow, S. M.; Kogan, T. P.; Venuti, M.; Gadek, T.; Harris, R. J.; Peers, D. H.; Mordenti, J.; Shak, S.; Ashkenazi, A. *Bioconjugate Chem.* **1994**, 5, 133.
13. A full account of cellular and bioassay studies will be published in due course. Preliminary data can be obtained from the corresponding author.
14. Rago, R. P.; Einstein, A.; Lush, R.; Tomasz, M. B.; Ko, Y. J.; Henner, W. D.; Bubley, G. J.; Merica, E. A.; Garg, V.; Ette, E.; Harding, M. W.; Dalton, W. S. *Cancer Chemother. Pharmacol.* **2003**, 51, 297.